University of Nebraska - Lincoln DigitalCommons@University of Nebraska - Lincoln

Publications, Agencies and Staff of the U.S. Department of Commerce

U.S. Department of Commerce

2007

Effective population size of steelhead trout: influence of variance in reproductive success, hatchery programs, and genetic compensation between life-history forms

Hitoshi Araki Oregon State University Corvallis, arakih@science.oregonstate.edu

Robin Waples NOAA, robin.waples@noaa.gov

William Ardren Oregon State University, William_Ardren@fws.gov

Becky Cooper Oregon State University, rdcooper@pdx.edu

Michael S. Blouin Oregon State University, blouinm@science.oregonstate.edu

Follow this and additional works at: http://digitalcommons.unl.edu/usdeptcommercepub

Araki, Hitoshi; Waples, Robin; Ardren, William; Cooper, Becky; and Blouin, Michael S., "Effective population size of steelhead trout: influence of variance in reproductive success, hatchery programs, and genetic compensation between life-history forms" (2007). *Publications, Agencies and Staff of the U.S. Department of Commerce.* 465. http://digitalcommons.unl.edu/usdeptcommercepub/465

This Article is brought to you for free and open access by the U.S. Department of Commerce at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Publications, Agencies and Staff of the U.S. Department of Commerce by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Effective population size of steelhead trout: influence of variance in reproductive success, hatchery programs, and genetic compensation between life-history forms

HITOSHI ARAKI,* ROBIN S. WAPLES,† WILLIAM R. ARDREN,*‡ BECKY COOPER* and MICHAEL S. BLOUIN*

*Department of Zoology, Oregon State University, 3029 Cordley Hall, Corvallis, Oregon 97331, USA, +Northwest Fisheries Science Center, 2725 Montlake Blvd. East, Seattle, Washington 98112, USA

Abstract

The effective population size is influenced by many biological factors in natural populations. To evaluate their relative importance, we estimated the effective number of breeders per year (N_h) and effective population size per generation (N_a) in anadromous steelhead trout (Oncorhynchus mykiss) in the Hood River, Oregon (USA). Using demographic data and genetic parentage analysis on an almost complete sample of all adults that returned to the river over 15 years (> 15 000 individuals), we estimated $N_{\rm h}$ for 13 run years and N_{e} for three entire generations. The results are as follows: (i) the ratio of N_{e} to the estimated census population size (N) was 0.17–0.40, with large variance in reproductive success among individuals being the primary cause of the reduction in N_e/N_i (ii) fish from a traditional hatchery program (H_{trad} : nonlocal, multiple generations in a hatchery) had negative effects on N_{h} , not only by reducing mean reproductive success but also by increasing variance in reproductive success among breeding parents, whereas no sign of such effects was found in fish from supplementation hatchery programs (H_{supp} : local, single generation in a hatchery); and (iii) N_b was relatively stable among run years, despite the widely fluctuating annual run sizes of anadromous adults. We found high levels of reproductive contribution of nonanadromous parents to anadromous offspring when anadromous run size is small, suggesting a genetic compensation between life-history forms (anadromous and nonanadromous). This is the first study showing that reproductive interaction between different life-history forms can buffer the genetic impact of fluctuating census size on N_{e} .

Keywords: effective population size, genetic compensation, microsatellite, *Oncorhynchus mykiss*, parentage analysis, salmonid

Received 14 July 2006; revision accepted 23 October 2006

Introduction

The effective population size (N_e) is one of the most important parameters in evolutionary and conservation biology, not only because N_e determines the degree of genetic drift and the effectiveness of natural selection, but also because it affects population viability (Frankham *et al.*)

Correspondence: Hitoshi Araki, Fax: 1-541-737-0501; E-mail: arakih@science.oregonstate.edu

‡Present address: Conservation Genetics Laboratory, Abernathy Fish Technology Center, U.S. Fish and Wildlife Service, 1440 Abernathy Creek Road, Longview, Washington 98632, USA 2002; Hedrick 2005a). However, this parameter is also one of the most difficult to estimate in natural populations. Many factors influence N_e including sex ratio, variance in family size among individuals, fluctuations in population size, and age structure (Crow & Kimura 1970). As a consequence, N_e is often much less than the census size (N) in many species (e.g. Husband & Barrett 1992; Scribner *et al.* 1997; Turner *et al.* 2002). Because N is generally much easier to measure than N_e , there has been much interest in predicting N_e/N ratios, particularly for conservation applications (Frankham 1995a, 2002). However, how best to measure N_e/N ratios, and whether they are constrained within a narrow range in nature or can take extremely low values

for some species, remain open questions (Nunney 1993, 1995; Frankham 1995b; Hauser *et al.* 2002; Kalinowski & Waples 2002; Hedrick 2005b). In addition, the factors that can reduce N_e/N ratios usually act simultaneously, which makes it difficult to distinguish their individual effects. As a consequence, few empirical studies have addressed the relative importance of the different factors above in reducing N_e/N in natural populations (Frankham 1995b; Storz *et al.* 2002; Ardren & Kapuscinski 2003).

Two additional factors may affect N_e and N_e/N in practice. One is artificial propagation, in which natural populations are supplemented with cultured individuals. Artificial propagation is common for economically valuable species (Leber 2004), but many conservation programs also utilize artificial propagation to support natural populations of endangered species (Cuenco *et al.* 1993; Olney *et al.* 1994). These species now include many hundreds of taxa in both aquatic and terrestrial systems (Frankham 2002). Although the effects of artificial propagation on N_e in natural populations have been evaluated theoretically (Ryman & Laikre 1991; Waples & Do 1994; Wang & Ryman 2001; Duchesne & Bernatchez 2002), empirical data on this important topic are limited (Ballou & Foose 1996; Hedrick *et al.* 2000; Gautschi *et al.* 2003).

Another potential factor that can affect N_{e} and N_{e}/N is life history polymorphisms (e.g. behavioural polymorphisms such as resident and migratory forms, and alternative mating strategies such as dominant males vs. sneaker males). Different life-history forms can have different reproductive success in different circumstances, and interbreeding between life-history forms might buffer fluctuations in N_e in some circumstances. For example, if abundance of one life-history form fluctuates widely, while that of the other form is relatively stable, then overall N_e will be stabilized through time by interbreeding between forms. Similarly, if the sex ratio of one form is heavily skewed the overall sex ratio (and hence N_{e}) can be balanced by interbreeding between life-history forms. This topic is just beginning to be studied in detail, but already it is clear that the relative contributions of different life-history forms can have important consequences for effective population size (Martinez et al. 2000; Garcia-Vazquez et al. 2001). Parallel situations occur in many marine fish species that have sex reversal (Muñoz & Warner 2003) and in a variety of species (from insects to mammals) that have dominant and sneaker males (Wade & Shuster 2004).

Salmonid species provide a good model system for evaluating N_e and N_e/N ratios. Salmonids are of considerable economic, social, and cultural interest, and they spawn at predictable times in discrete freshwater populations. As a result, these species have been well studied and census population sizes can often be estimated very accurately. Anadromous Pacific salmonids (*Oncorhynchus* spp.) are also semelparous, or nearly so, which facilitates conversion of estimates of the effective number of breeders per year (N_b) into effective size per generation (N_e) (Waples 2002a). These life history traits — semelparity with variable age at maturity — are shared by a number of other taxa, including many monocarpic plants and crustaceans with diapausing eggs (Waples 2006a). Genetic data have long been used in the conservation and management of salmonid populations, so a large number of markers are available for estimating effective size and N_e/N ratios using genetic data (Hedrick *et al.* 2000; Heath *et al.* 2002; Ardren & Kapuscinski 2003; Waples 2004).

Oncorhynchus mykiss has two distinct life-history forms: steelhead trout, which are anadromous, and resident rainbow trout, which stay in freshwater throughout the life cycle (hence nonanadromous). Despite frequent interbreeding between anadromous and nonanadromous fish in natural populations (Zimmerman & Reeves 2000; Salmon Recovery Science Review Panel 2004), the influence of this interbreeding on N_{ρ} has never been studied. In a previous study, we found large fluctuations in the numbers of returning adults among years, low reproductive success of adult fish from a traditional hatchery program in the wild, and a large reproductive contribution of nonanadromous fish to anadromous populations in steelhead populations in the Hood River, Oregon (Araki et al. 2006). Thus, this system is ideal for evaluating the effect of interbreeding between different life-history forms on N_h and N_e .

In this study, we use demographic data and microsatellitebased parentage assignments to estimate N_h and N_e in two steelhead populations (summer run and winter run) in the Hood River. Our data set includes more than 15 000 samples, representing almost all (>97%) anadromous adults that returned to spawn in the river between 1991 and 2003. We investigate 13 run years of parents and their offspring (that returned to the river as adults in subsequent run years), which covers one full generation of summer run and two full generations of winter run. We address: (i) N_{e} *N* ratios in these populations and the influence of various factors that affect N_b and N_e (sex ratio, variance in reproductive success among individuals, temporal variation in reproductive success) on N_{ρ}/N ; (ii) the effects of hatchery propagations on N_h and N_{ρ} ; and (iii) temporal fluctuation (or stability) of N_h and its cause in the natural populations. This is the first study to estimate N_{e} directly from almost complete demographic data for entire generations of anadromous populations of a salmonid. Although we do not have nonanadromous samples, the almost complete sampling from anadromous steelhead and the parentage analysis allow us to infer the relative contribution of nonanadromous parents to the anadromous populations.

Materials and methods

See Table 1 for an explanation of notation.

Table 1 Notation used in this paper

N _e	Effective population size per generation
N _b	Effective number of breeders per run year
N _{anad}	Number of anadromous fish passed above the dam each run year; this includes both hatchery-born and wild-born fish
Ν	Total number of potential parents, including anadromous and nonanadromous parents
Ń	Estimate of <i>N</i> based on equations 1 and 2
k	Mean reproductive success (family size) among parents
V_k	Variance in reproductive success (family size) among parents
V_k/\bar{k}	Index of variability (= variance in reproductive success divided by mean reproductive success)
	'Adjusted V_k/\bar{k}' means using \bar{k} adjusted for the bias that results from parentage assignment errors (via equation 3)
	'Scaled V_k/\bar{k}' means scaled to a stable population size of $\bar{k} = 2$ (also via equation 3)
	This scaling allows one to compare the index of variability among different datasets
$H_{\rm trad}$	Fish from the traditional hatchery program using nonlocal brood stock that reared multiple generations in hatcheries
H_{supp}	Fish from the supplementation hatchery program using local, wild brood stock

Study populations

The Hood River basin is a tributary of the Columbia River in the Northwest United States. Since 1991 almost every adult steelhead returning to spawn in this river has been catalogued, measured, and had scale and fin-snip samples taken (for DNA analysis) at the Powerdale Dam fish trap by staff of the Oregon Department of Fish and Wildlife (ODFW). This dam is located 4.0 river miles from the mouth of the Hood River. Steelhead only spawn above the dam, which is a complete upstream barrier to migration for all salmonids. The year in which each returning adult was born (brood year) was determined via scale reading, and the average generation time (birth-year of parents to birthyear of offspring) was estimated as 5.2 years for summer runs and 4.3 years for winter runs. Although there is some overlap in the run timing of the two populations (summer run: April to October, winter run: January to June Kostow 2004), summer run and winter run fish are usually quite distinguishable because they only overlap in run timing for a brief period, and during this time winter run are sexually mature and summer run are not. Both runs breed in the spring, but in different forks of the river. Fewer hybridization events between populations were identified in our parentage analysis than the number of matches expected by chance alone given our empirical estimates of assignment error rates (i.e. type-B error rates, see below), indicating there is little or no hybridization between the two run populations. Indeed, microsatellite-based F_{ST} between run populations was 0.009, nearly 10 times higher than temporal F among run years within populations (F = 0.001) (Hedrick 2005a). Steelhead is an iteroparous species (repeated reproductive bouts possible), but we treated all the fish as semelparous (only one reproductive bout in a lifetime) because the proportion of repeat spawners is very low in this river (< 5%).

In a previous study on the Hood River (Araki *et al.* 2006), we found that a large fraction of anadromous fish did

not have one or both parents among the anadromous spawners from the year in which they were born. Here we call these missing parents 'nonanadromous parents', because we have almost complete samples from anadromous parents for the run years we examined. We believe that the majority of the missing parents are resident trout, which are common in the Hood River (Olsen 2003). Precocious parr (presmolts that mate before going to sea) might also be involved, but they have not been widely reported in steelhead. Strays from other rivers could also explain the missing parents. However, we believe that strays are not a major source of the missing parents in this river for the following reasons: (i) a large proportion of offspring had at least one parent from the Hood River (Araki et al. 2006); (ii) offspring with both parents missing (potential strays) are genetically indistinguishable to offspring whose parents are in the sample (unpublished data); and (iii) according to the hatchery-born fish identification, the straying rate from other rivers to the Hood River is low (2%-3%; E. Olsen, personal communication).

For parents, the run year is the year in which adult fish begin arriving at the river to spawn, and is the calendar year before they actually spawn. Hence fish from the 1995 run year began arriving in 1995 and spawned in the spring of 1996 (which is the brood year of their offspring). In this study, we investigated five consecutive run-year samples of summer run (run years 1994-98; hereafter Su94 to Su98) that roughly cover one complete generation in the summerrun population, and eight consecutive run-year samples of winter run (run years 1991-98; hereafter Wi91 to Wi98) that roughly cover two consecutive generations in the winterrun population. The adult samples that returned to the river between run year 1994 and 2003 for winter run and between run year 1998-2003 for summer run were used as offspring samples, based on their brood years. Because our samples cover more than one generation, some samples were used both as offspring samples (in earlier sample sets) and as parent samples (in later sample sets). The total

Run year	Parents		Offspring		
	N _{anad}	%Wild	%Male	N _{Offspring}	$N_{ m Offspring}/N_{ m anad}$
Summer run (Su	ı)				
Su94	1831	10.3	40.8	198	0.11
Su95	650	20.3	39.7	208	0.32
Su96	1486	12.2	37.5	588	0.40
Su97	529	15.7	33.5	456	0.86
Su98	136	97.1	27.9	240	1.76
Su total	4632	15.5	38.4	1690	0.36
Winter run (Wi)					
Wi91	1008	71.0	46.3	271	0.27
Wi92	412	98.8	38.6	303	0.74
Wi93	384	99.5	32.6	212	0.55
Wi94	208	97.1	50.0	296	1.42
Wi95	461	59.9	47.5	1233	2.67
Wi96	572	50.7	40.4	983	1.72
Wi97	422	52.8	37.2	879	2.08
Wi98	517	57.4	42.2	612	1.18
Wi total	3984	70.1	42.2	4789	1.20

 N_{anad} represents a number of anadromous steelhead adults returned to the Hood River and passed above the dam (complete upstream barrier to the spawning grounds), and $N_{\text{Offspring}}$ represents a number of anadromous steelhead adults that were born in the wild in the year when corresponding parental adults spawned, and that returned to the Hood River in the subsequent run years (mostly as 2–6 years old). We identified the brood years of returning offspring by ageing their scales. %Wild and %Male represent percentages of wild-born fish and of males in each run year, respectively.

In Su94–97 and Wi91, traditional hatchery programs were operated individually for summer run and winter run, and all the returned fish were allowed to pass above the dam. In Su98 and Wi92–94, almost none of the returned fish born in the traditional hatcheries were allowed to pass above the dam, so that only wild-born fish could reproduce in these run years. In Wi95–98, both wild-born fish and fish from a supplementation hatchery had returned to the river, and the numbers of hatchery-born fish passed above the dam were controlled to be < 50% of all the fish allowed to pass.

numbers of individuals used in this study were 4561 for summer-run parents, 2217 for summer-run offspring, 3945 for winter-run parents, and 5991 for winter-run offspring. Note that the parental samples include both wild-born and hatchery-born fish, whereas the offspring samples include only wild-born fish (Table 2).

Hatchery programs in the Hood River

Traditional hatchery stocks, H_{trad} , are produced using nonlocal brood stock that has spent multiple generations in hatcheries. Supplementation hatchery stocks, $H_{supp'}$ are produced using local, wild fish as broodstock. The Hood River has been stocked by the ODFW since 1960s with winter-run and summer-run H_{trad} . The winter-run H_{trad} was phased out of the Hood River in the early 1990s, while the summer-run H_{trad} was phased out in the late 1990s. The supplementation programme of steelhead in this river started in 1991 for the winter-run populations, and in 1997 for the summer-run populations. The winter-run H_{supp} began breeding in the river in appreciable numbers in 1995, and the summer-run H_{supp} in 2001. Details of the supplementation program are described in Olsen (2003) and Araki *et al.* (2006).

In Su94–97 and Wi91, all the returning fish were allowed to pass above the dam, including high proportions of $H_{\rm trad}$ (Table 2). As part of an effort to phase out $H_{\rm trad}$, almost none of the traditional hatchery-born fish were allowed to pass above the dam in Su98 and Wi92–94. Thus, almost all of the winter-run fish that reproduced in run years Wi92–94 were wild-born. In Wi95–98, appreciable numbers of $H_{\rm supp}$ started to return, and the number of $H_{\rm supp}$ passed above the dam was controlled to be < 50% of all the fish allowed to pass, to avoid a predominance of hatchery-born spawners in this river (and hence in the gene pool of the populations).

Microsatellite loci and parentage analysis

Genotypes at eight microsatellite loci (Omy1001, Omy1011, Omy1191, Omy77, One108, One2, Ssa407, and Str2, see Araki *et al.* 2006 for details) were identified for these samples. We followed a standard Chelex protocol for DNA extraction and amplification (see Nelson *et al.* 1998), with

minor modifications (50–55 °C of annealing temperature). The genotype scoring was done on an ABI 3100 capillary electrophoresis system (Applied Biosystems, California). We successfully genotyped 97.6% of the fish that returned to the river as adults (Table 2), at up to eight loci. The average number of genotype-determined loci was 7.77 per sample. Part of the genotype data was used in Araki *et al.* (2006).

We used an exclusion method to make parentage assignments (Araki *et al.* 2006). Putative parent-offspring pairs sharing at least six loci (some fish had missing data) and having no mismatching loci were assigned to parentoffspring pairs using the CERVUS program (Marshall *et al.* 1998). In the few cases that more than one candidate parent shared no mismatch to an offspring, the parent with the highest LOD score was assigned. Total exclusionary powers were at least 0.9996 in all cases.

Estimating the whole (census) population size

To estimate the ratio N_e/N , the census population size was estimated based on the run size (census size of returned anadromous adults, N_{anad}). Because our samples were collected at the complete upstream barrier to steelhead, parental population size listed in Table 2 represents virtually all the anadromous fish potentially reaching the spawning grounds. As mentioned above, however, noticeable numbers of parents turned out to be missing from the dataset in the parentage assignment (Araki *et al.* 2006), suggesting that nonanadromous parents produced a substantial number of anadromous offspring.

To take the nonanadromous parents into consideration for estimating parental $N(\hat{N})$, we calculated \hat{N} for each sex separately as follows:

$$\hat{N} = \frac{N_{\text{anad}}}{\hat{P}_{\text{sampled}}} \tag{eqn 1}$$

$$\hat{P}_{\text{sampled}} = \frac{1}{(1-\hat{a})(1-\hat{b})} \left(\frac{N_{\text{assigned}}}{N_{\text{offspring}}} - \hat{b} \right)$$
(eqn 2)

where $\hat{P}_{sampled}$ is the proportion of offspring whose parent is in the sample (either father or mother), $N_{offspring}$ and $N_{assigned}$ are numbers of returned offspring and assigned offspring, and \hat{a} and \hat{b} are the type-A and type-B error rates, respectively (Araki & Blouin 2005; Araki *et al.* 2006). The type-A error rate is the rate for failing to assign a true parent when that parent is in the sample, and the type-B error rate is the rate at which nonparents are incorrectly assigned to offspring. In this study, we used the error rates previously estimated based on the same dataset and the same criteria of parentage assignment $\hat{a} = 15.6\%$ and $\hat{b} = 1.75\%$ for male parents and $\hat{a} = 5.7\%$ and $\hat{b} = 1.97\%$ for female parents, which were estimated using known parentoffspring pairs of brood stock and their hatchery-born offspring from run years Wi93 and Wi94 in Araki *et al.* (2006)]. After obtaining \hat{N} for each sex, \hat{N} for each run year was calculated as:

$$\hat{N}_{[\text{run-year}]} = \hat{N}_{[\text{male}]} + \hat{N}_{[\text{female}]}$$

to estimate \hat{N}_b/\hat{N} . For \hat{N}_e/\hat{N} , \hat{N} in each generation was calculated simply by summing up $\hat{N}_{[run-year]}$ within each generation.

An important assumption here is that the ratio of the numbers of nonsampled 'parents' to sampled 'parents' is the same as the ratio of the numbers of 'offspring' whose parents were not sampled to those sampled (i.e. $[1 - P_{sampled}]/P_{sampled}$). This assumption holds only when the mean reproductive success (\bar{k}) among nonsampled parents is equal to \bar{k} among sampled parents, and may be unrealistic considering differences in phenotypes and reproductive strategies between anadromous and nonanadromous fish. To confirm the robustness of our conclusions against a violation of this assumption, a simple assumption of equal numbers of sampled and nonsampled parents ($\hat{N} = 2N_{anad}$) was also made and the results were compared.

Angling for hatchery steelhead trout was allowed above the dam in a limited area during 1993–98. We used the ODFW upper-bound estimates of the harvest rate on hatchery fish in the Hood River of 25% for Su94–97, 5.0% for Wi95, 20% for Wi96–97 (E. Olsen, personal communication) to estimate the minimum number of potential spawners in those years.

Demographic estimate of N_e

Based on the parentage assignments, the mean (\bar{k}) and variance (V_k) of reproductive success among parents were estimated for each sex in each parental population. Because parentage assignment errors can affect the estimation of \bar{k} and $V_{k'}$ we first adjusted the observed \bar{k} using the method of Araki & Blouin (2005). An unbiased estimate of \bar{k} ($\bar{k}_{[obs]}$) was obtained by correcting the bias caused by the assignment errors (*a* and *b* above) using equation 13 in Araki & Blouin (2005). Because we could not directly estimate an unbiased $V_{k'}$ we adjusted the observed index of variability ($V_{k[obs]}/\bar{k}_{[obs]}$) to obtain an adjusted value by:

$$\left[\frac{V_{k[\text{adj}]}}{\bar{k}_{[\text{adj}]}}\right] \approx 1 + \frac{\bar{k}_{[\text{adj}]}}{\bar{k}_{[\text{obs}]}} \left(\frac{V_{k[\text{obs}]}}{\bar{k}_{[\text{obs}]}} - 1\right)$$
(eqn 3)

where $V_{k[adj]}$ is the adjusted variance in reproductive success (Crow & Morton 1955; Waples 2002b). While this approximation was originally applied to adjust the index of variability between different life stages (equation 14 in Crow & Morton 1955), we applied it to adjust the index for $\bar{k}_{[adj]}$ (unbiased estimate of \bar{k}) here.

After adjusting V_k/\bar{k} for the assignment bias, we estimated the inbreeding N_b for each sex $(N_{b[\text{female}]} \text{ or } N_{b[\text{male}]})$ using:

$$N_b \approx \frac{\bar{k}N - 2}{\bar{k} - 1 + V_k/\bar{k}} \tag{eqn 4}$$

(Crow & Kimura 1970; Caballero 1994), and the estimate of N_b in run year *i* (N_{blil}) was obtained using:

$$N_{b[i]} = \frac{4(N_{b[\text{female}]} \times N_{b[\text{male}]})}{(N_{b[\text{female}]} + N_{b[\text{male}]})}$$
(eqn 5)

Finally, N_e for one generation was estimated using:

$$N_{e} = \frac{1}{\sum (X_{i}^{2}/N_{b[i]})}$$
 (eqn 6)

where X_i is the proportional contribution of breeders in run year *i* to the next generation (Waples 2002a, 2006a). For simplicity, five years/generation for summer run and four years/generation for winter run were assumed (see above). Using the samples from five summer-run years and eight winter-run years, we therefore obtained N_e in one complete generation of summer run and in two complete generations of winter run in this study.

For general comparisons of the index of variability, equation 3 was further used for scaling to $\bar{k} = 2$ (by replacing $\bar{k}_{[adj]}$ with 2). We call it the scaled index of variability, contrasting to the adjusted index of variability above (Table 1). Note that inbreeding N_b based on the adjusted index of variability is almost identical to variance N_b based on the scaled index of variability (Waples 2002b).

Evaluating reduction factors of N_e

To examine the importance of sex ratio and two components of variance in reproductive success (variance among individuals within years and variance in \bar{k} across years within a generation) on reducing N_e/N , we calculated N_e/N independently for each of these factors.

To evaluate the effect of sex ratio, we first calculated the proportion of male parents (P_{male}) by adding up all the numbers of males and females in a generation (Table 2). Equation 5 provides an estimate of N_e/N reflecting only the sex ratio as $4P_{male}(1 - P_{male})$. The reduction rate was then calculated as $(1 - N_e/N)$. In this case, $N_e/N = 1$ (reduction rate = 0) when the sex ratio is 1 : 1.

Likewise, equation 4 can provide an estimate of N_e/N affected only by the variance in reproductive success among individuals. To evaluate this effect, we first calculated an overall V_k/\bar{k} as the geometric mean (GM) of the scaled V_k/\bar{k} ($\bar{k} = 2$) over run year and sex, ignoring the sex ratio bias and the difference in N among run years. Equation 4 was then used for estimating N_e (instead of N_b in equation 4), by replacing N in each run year with \hat{N}_{total} (total \hat{N} in a generation, Table 3). The reduction rate by this factor is given as: $(1 - \hat{N}_e / \hat{N}_{\text{total}})$

Here we ignore prespawning mortalities, which are estimated as 10% for summer run and 5% for winter run (E. Olsen, personal communication), because this factor affects the expectations of N_e/N only slightly, as is apparent from equation 4. In this case, $N_e = (N - 1)$ when the scaled $V_k/\bar{k} = 1$.

Finally, we evaluated the effect of variance in \bar{k} across years within a generation (see Waples 2006a). Equation 6 can provide an estimate of N_e reflecting only this factor, by replacing $N_{b[i]}$ with \hat{N} in each run year (Table 3). Again, the reduction rate was given as $(1 - \hat{N}_e / \hat{N}_{total})$, but in this case $N_e / N = 1$ when \bar{k} is constant among run years within a generation (\bar{k} = total number of offspring in a generation divided by \hat{N}_{total} in this case).

Distribution of family size

For some example years, we compared the observed distribution of family size with that expected under the Poisson and negative binomial distributions. The Poisson distribution characterizes an ideal population with $N_h = N$. The negative binomial distribution represents an overdispersed Poisson distribution with $V_k > \bar{k}$ and $N_h < N$. We used a two-step process to generate a negative binomial distribution with the same mean and variance in family size as the observed distribution (Anderson 2001; Waples 2006b). First, a random sample was taken from a gamma distribution with the appropriate shape and scale parameters. Next, this gamma random number was used as the parameter for a Poisson distribution, from which a random number was drawn to represent the family size of one individual. This process was repeated 200 000 times to generate a smooth, negative binomial distribution of expected family size. Goodness of fit to the observed distribution was evaluated by a chi-square test with 27 classes (family sizes (k) = 0–25, and a combined class for k > 25) and 24 degrees of freedom.

This two-stage process departs from the standard Wright-Fisher ideal population in having both a random and a directed process (Anderson 2001). In an ideal population, each individual contributes equally to an infinite gamete pool, from which the next generation is randomly drawn. In the two-step model, the *i*th individual contributes G_i gene copies to the gamete pool, with G_i following the gamma distribution. When the next generation is randomly drawn from this gamete pool, the result is a larger variance in family size than occurs in an ideal population.

Results

Variance in reproductive success

Estimated indexes of variability (V_k/\bar{k}) are shown for each sex in Table 3. The V_k/\bar{k} scaled to $\bar{k} = 2$ was 5.8–17.3 for

Run year	Ń	Adjusted V_k/\bar{k}		Scaled V_k/\bar{k} ($\bar{k} = 2$)				
		Male	Female	Male	Female	\hat{N}_b	\hat{N}_b/\hat{N}	$\hat{N}_b/2N_{ m anad}$
Summer run (Su)							
Su94	2506	1.5	1.4	8.4	7.6	547.4	0.22	0.15
Su95	1518	1.2	2.7	10.3	17.3	204.2	0.13	0.16
Su96	1902	2.7	2.2	6.7	6.7	478.3	0.25	0.16
Su97	1500	3.6	3.0	9.4	9.7	278.4	0.19	0.26
Su98	927	1.0	2.3	6.0	5.8	230.6	0.25	0.85
Su total	8354	2.0*	2.3*	8.0*	8.7*	1425.9**	0.17	0.15
Winter run ge	neration 1 (Wi	i-1)						
Wi91	1781	2.0	1.7	7.7	4.9	491.6	0.28	0.24
Wi92	710	3.5	2.1	6.5	3.6	228.6	0.32	0.28
Wi93	760	2.2	1.6	4.4	3.0	314.3	0.41	0.41
Wi94	512	2.3	2.4	3.9	2.4	233.8	0.46	0.56
Wi-1 total	3762	2.4*	1.9*	5.4*	3.3*	1095.5**	0.29	0.27
Winter run ge	neration 2 (Wi	i-2)						
Wi95	700	6.5	4.7	4.2	2.7	316.7	0.45	0.34
Wi96	921	3.5	2.1	3.3	2.1	501.1	0.54	0.44
Wi97	868	2.8	2.9	3.0	2.6	452.1	0.52	0.54
Wi98	1395	2.8	1.9	5.2	3.0	541.8	0.39	0.52
Wi-2 total	3884	3.6*	2.7*	3.8*	2.6*	1505.3**	0.40	0.38

Table 3 Demographic estimate of $N_{b'}$, $N_{e'}$ and $N_{e'}/N$

 \hat{N} represents an estimate of census population size in a parental population, based on equation 1. V_k/\bar{k} is the index of variability, estimated based on the parentage analysis. Adjusted V_k/\bar{k} was estimated only by adjusting parentage assignment errors, whereas scaled V_k/\bar{k} was weighted by $\bar{k} = 2$ (see Table 1 for precise definitions). \hat{N}_b was estimated by the demographic method using equation 5.

*Geometric mean among run years within a generation.

** N_e per generation estimated based on equation 6.

summer run (GM = 8.0 for male, 8.7 for female) and 2.1– 7.7 for winter run (GM = 4.5 for male, 2.9 for female), suggesting larger variance in reproductive success in both sexes and both populations compared with that expected for a Poisson distribution (V_k/\bar{k} = 1). Indeed, some individuals evidently left > 20 offspring that returned to the river successfully, despite little or no reproductive success for most parents (Fig. 1 for example of Wi95). Males had significantly higher scaled V_k/\bar{k} than females in all run years of winter run (*F*-test after square-root transformation, P = 0.000-0.035, combined P < 0.001), whereas the difference between sex was not significant in summer run except Su91 (P = 0.002-1.0, combined P = 0.039).

The example shown in Fig. 1 illustrates the dramatic difference between the observed distribution of family size and that expected from a Poisson distribution. In contrast, the fit to the negative binomial distribution was very good for the males (chi square = 19.7; d.f. = 24; P > 0.5) and not quite as good for the females (chi square = 44.0; d.f. = 24; 0.005 < P < 0.01) but still dramatically better than the fit to the Poisson distribution. This result suggests that reproductive success in *Oncorhynchus mykiss* can be modelled as a two-stage process resulting in an overdispersed Poisson distribution, at least in some cases. The explanation for the slightly poorer fit for the female example (Fig. 1b) is not clear, but factors that might be involved include a high rate of infertility and/or inaccurate estimates of harvest mortality (both of which could inflate the number of families leaving no offspring) and chance events.

Demographic estimate of effective population size

Estimated N_b was very similar in the two run populations (204-547 in summer run and 229-542 in winter run, Table 3). These estimates were also relatively stable among run years, despite the highly variable census size of anadromous adults (N_{anad}) in these run years (Fig. 2). The unbiased estimates of coefficient of variation (CV, Sokal & Rohlf 1995) in \hat{N}_b were lower than in N_{anad} in both populations $(CV[\hat{N}_b] = 46.8 < CV[N_{anad}] = 80.0$ in summer run, $CV[\hat{N}_b] = 33.7 < CV[N_{anad}] = 48.1$ in winter run), indicating stable \hat{N}_{b} relative to $N_{\rm anad}$ \hat{N}_{e} (per generation) was 1096– 1505, and overall \hat{N}_{ρ}/\hat{N} ratio was 0.17–0.40 (Table 3). These ratios were very similar to $\hat{N}_e/(2N_{anad})$, suggesting that the results were not biased by errors in estimating the census population sizes. One exception among run year was \hat{N}_{h} / $(2N_{anad}) = 0.85$ in Su98, which was clearly different from \hat{N}_{b}/\hat{N} in the same run year ($\hat{N}_{b}/\hat{N} = 0.25$, see below).



Fig. 1 Example histograms for number of offspring produced by each parent in Wi95. Male (a) and female (b), and observed number from parentage assignments (open bar), expected number from a negative binomial (open circle), and expected number from Poisson distribution (closed square) were drawn separately (see Materials and methods). $\bar{k}_{\rm [obs]}$ was 3.3 for male and 4.4 for female, and $V_{k\rm [obs]}$ was 18.1 for male and 18.4 for female in Wi95. Among male, for example, 45 males left no returning offspring, whereas one male left 21 offspring in Wi95. Note that the observed numbers are considered as minimum estimates, because assignment-error biased these estimates downward and because we used the ODFW upper-bound estimates of the harvest rate.

Given the observed sex ratio in anadromous parents (Table 2), census sizes of anadromous adults in each run year, and the estimated (scaled) index of variability (Table 3), we evaluated how much reduction of $N_{e'}$ relative to N, was caused by these factors (Table 4). The reduction rates $(1 - N_e/N)$ were estimated as only 2.3%–5.4% by une-



Fig. 2 Estimated effective number of breeders (N_b) and run sizes (N_{anad}) in two populations of steelhead in the Hood River. The summer run population is shown by circles (open circle for N_b , closed circle for N_{anad}) and the winter run population is shown by squares (open for N_b , closed for N_{anad}). H_{trad} and W: run year with a high proportion of traditional hatchery-born fish, W only: run year with virtually wild fish only, and H_{supp} and W: run year with up to 50% of supplementation hatchery-born fish. The unbiased estimates of coefficient of variation (Sokal & Rohlf 1995) were 80.0 for N_{anad} and 46.8 for \hat{N}_b in summer run, 48.1 for N_{anad} and 33.7 for \hat{N}_b in winter run, representing stable \hat{N}_b relative to N_{anad} .

Table 4 Effects of sex ratio and variance in reproductive success on N_e/N

Generation	Su	Wi-1	Wi-2
Reduction rate by each factor (%)			
unequal sex ratio	5.4	2.3	2.7
variance in reproductive success			
variance in \bar{k} among run years	19.9	22.4	19.2
large V_k/\bar{k} among individuals	78.5	61.9	51.7
Total expected N_e/N	0.16	0.29	0.38
Observed \hat{N}_b/\hat{N}^*	0.17	0.29	0.40

Total expected N_e/N was calculated by multiplying [1 – reduction rate] for the three reduction factors within each generation. *Data from Table 3.

qual sex ratio, but variance in reproductive success had a huge impact on N_e (reduction rate = 52%–79% by variance in reproductive success among individuals and 19%–22% by variance in \bar{k} across years within a generation), suggesting that high variance in reproductive success is a primary cause of $\hat{N}_e/\hat{N} < 1$ in these populations.

Assuming an independence of these factors, we obtained an expected N_e/N that was very similar to the \hat{N}_e/\hat{N} (Table 4), suggesting that the assumption of independence was reasonable. However, it is noteworthy that these estimates are based on the information only from anadromous parents. The parental population as a whole (including nonanadromous fish) may have different reduction rates and hence different N_e/N .

Effects of hatchery-born fish on N_b

Notably poor reproductive success is evident in the H_{trad} (Araki et al. 2006), which is the primary reason for the poor recruitment in Su94–97 and Wi91 ($N_{Offspring}/N_{anad}$, Table 2). Comparing with scaled V_k/\bar{k} in run years with mostly wild-born fish (Su98, Wi92–94), the larger scaled V_k/\bar{k} in Su94-97 and Wi91 (Table 3) indicates that the poor performance of H_{trad} led to high variance in reproductive success in the natural spawners as a whole (which is a primary factor causing small N_{e}/N). Separate estimates of the scaled V_k/\bar{k} for the wild-born parents and for the hatchery-born parents in the same run years support this view (Fig. 3). The scaled V_k/\bar{k} varied substantially between the wildborn and $H_{\rm trad}$ parents (e.g. in Su94 scaled V_k/\bar{k} was 4.2 in wild-born males and 8.2 in H_{trad} males). The geometric mean of the scaled V_k/\bar{k} in Su94–97 was 5.7 in wild-born male, 6.9 in H_{trad} male, 6.7 in wild-born female, and 8.3 in $H_{\rm trad}$ female. Interestingly however, the scaled V_k/\bar{k} were not always higher in $H_{\rm trad}$ parents than in wild-born parents in some run years. In Su95, for example, wild-born parents had exceptionally high V_k/\bar{k} in both sexes (Fig. 3). The reason why the scaled V_k/\bar{k} in wild-born parents in Su94– 97 are more variable than in the other run years is unclear (see Discussion).

The scaled V_k/\bar{k} in H_{supp} parents, on the other hand, were close to those in the wild-born parents and to their overall V_k/\bar{k} in run years Wi95–98 (except Wi98 female, Fig. 3). The geometric mean of the scaled V_k/\bar{k} was 3.5 in wild-born male, 3.7 in H_{supp} male, 2.4 in wild-born female, and 2.8 in H_{supp} female in Wi95–98. The scaled V_k/\bar{k} in Wi95–98 (with H_{supp}) were also similar to those in Wi92–94 (almost no hatchery fish returned), suggesting little effect by H_{supp} on the index of variability in the natural spawners.

Genetic compensation

The stable N_b among run years relative to highly variable run sizes of anadromous adults (Fig. 2) indicates that some kind of compensation may have occurred. Indeed, Ardren & Kapuscinski (2003) found a density-dependent N_b/N in a steelhead population, suggesting 'genetic compensation' in which reduction in N_b is buffered when N is small, presumably owing to reduction in competition for mates or spawning sites ('genetic compensation *within anadromous*



Fig. 3 The scaled index of variability (the scaled V_k/\bar{k}) among wild-born parents (white bar) and hatchery-born parents (dark bar) in run years. H_{trad} and W: run year with a high proportion of traditional hatchery-born fish, W only: run year with virtually wild fish only, and H_{supp} and W: run year with up to 50% of supplementation hatchery-born fish. Difference in the scaled $V_k/$ $ar{k}$ between hatchery and wild fish was tested using F-test, after square-root transformation to adjust the skewed distribution of the number of offspring per parent (**P < 0.01). Comparing results among different run years, disturbance in the index of variability (V_k/k) by traditional hatchery-born parents is evident, suggesting a negative effect of traditional hatchery programmes on the index of variability (and hence on N_h/N). No such effect by the supplementation hatchery-born fish is detected, although H_{supp} females have significantly large V_k/k relative to wild females that returned in the same year in Wi98 (P < 0.001).

steelhead population', hereafter). In this case, positive correlation between V_k/\bar{k} and anadromous run size is expected. However, we found no evidence for such a correlation between scaled V_k/\bar{k} and $N_{\rm anad}$ in the summer-run population (r = 0.03, P = 0.962 for male, r = -0.22, P = 0.727





Fig. 4 Relationship between variance in reproductive success and run size among run years. Summer run (a) and winter run (b), and male (solid line and circle) and female (dotted line and triangle). V_k/\bar{k} was scaled to $\bar{k} = 2$ (see Materials and methods). r = 0.03, P = 0.962 for summer-run male, r = -0.22, P = 0.727 for summer-run female, r = 0.70, P = 0.051 for winter-run male, and r = 0.78, P = 0.024 for winter-run female by two-tailed *t*-test. The marginal significances for the winter run become nonsignificant, however, if the Wi91 data (the rightmost points for both sexes in B) are removed (r = 0.09, P = 0.84 for male, r = 0.15, P = 0.75 for female. See Discussion).

for female by two-tailed *t*-test, Fig. 4a). The correlation between scaled V_k/\bar{k} and N_{anad} was positive and moderately significant in the winter-run population (r = 0.70, P = 0.051 for male, and r = 0.78, P = 0.024 for female, Fig. 4b), but was not significant if the Wi91 data are removed (the rightmost points for both sexes in Fig. 4b). In Wi91, V_k/\bar{k} was substantially increased by traditional hatchery-born

parents (Fig. 3). Overall therefore we found no evidence for compensation within anadromous steelhead population in the summer run and at best weak evidence for the winter run.

Another possible explanation for the stable N_h is 'genetic compensation between life-history forms'. As we mentioned above, the interbreeding between life-history forms (anadromous and nonanadromous fish, in this case) might stabilize N_h among run years in the face of fluctuating census sizes. Because we obtained almost complete samples from anadromous parents, offspring whose parents were missing from our data must be attributed to the reproductive contribution of nonanadromous parents. Note that the missing parents are not an artefact caused by parentage assignment errors, because we took assignment errors into consideration (see Materials and methods). We found negative correlations between N_{anad} and the proportion of offspring with one or both parents missing in both runs and both sexes (Fig. 5), and the result from the summer run population was statistically significant (r = -0.93, P = 0.021for male, r = -0.98, P = 0.004 for female in summer run, and r = -0.45, P = 0.27 for male, r = -0.14, P = 0.75 for female in winter run). Overall, the combined probability of both sexes in the two populations (Sokal & Rohlf 1995) was also statistically significant (P = 0.004). This relationship also explains why $\hat{N}_b/2N_{anad}$ was so different from \hat{N}_b/\hat{N} in Su98 (Table 3), because in this run year N_{anad} was the smallest and \hat{N} was 5.8 N_{anad} rather than $2N_{\text{anad}}$.

Discussion

The principle results that emerged from our analyses are as follows: (i) the steelhead populations in the Hood River showed a large variance in reproductive success, which is the primary cause of the reduction in N_e/N ; (ii) the traditional hatchery-born fish had negative effects on N_b , not only by decreasing the mean reproductive success of the natural population but also by increasing the variance in reproductive success; and (iii) both run populations showed relatively stable N_b among run years and a negative correlation between anadromous run size and inferred reproductive contribution of nonanadromous parents. These results suggest a genetic compensation between lifehistory forms in this species. These points and their general implications are discussed below.

Variance in reproductive success in salmonids

The geometric means of the scaled V_k/\bar{k} were 8.0 in summer-run males, 8.7 in summer-run females, 4.5 in winter-run males, and 2.9 in winter-run females (Table 3). Although the large V_k/\bar{k} in summer-run will be in part due to the influence of traditional hatchery fish (Fig. 3), the scaled V_k/\bar{k} was also large in Su98 (5.77–5.95), in which almost no hatchery fish returned. The only comparable



Fig. 5 Negative correlation between the proportion of offspring with at least one parent missing from the data ($P_{\text{Imissing parents}}$) and run size among run years. $P_{\text{Imissing parents}}$ was estimated for each sex using equation 2 and $P_{\text{Imissing parents}} = (1 - P_{\text{sampled}})$. Summer run (a) and winter run (b), and male (solid line and circle) and female (dotted line and triangle). r = -0.93, P = 0.021 for summer-run male, r = -0.98, P = 0.004 for summer-run female, r = -0.45, P = 0.267 for winter-run male, and r = -0.14, P = 0.747 for winter-run female by two-tailed *t*-test. Overall, the combined probability for the negative correlation was 0.004.

published data from a natural salmonid population are for Atlantic salmon (*Salmo salar* L. Garant *et al.* 2001), in which unscaled V_k/\bar{k} for juvenile samples was 8.6 for males and 8.9 for females. After scaling the *Salmo* data to $\bar{k} = 2$, however, the index of variability dropped to 2.05 for males and 1.94 for females — much lower than we found for the steelhead populations. Results for some experimental hatchery populations of Pacific salmon also show less variance in reproductive success than we report here (scaled $V_k/\bar{k} = 1.1-5.1$; see Waples 2004 for reviews). Although further studies are necessary before general conclusions are drawn, this result indicates that steelhead populations in the Hood River have unusually large variance in reproductive success among individuals, even within salmonids. It is also noteworthy that males had significantly higher scaled V_k/\bar{k} than females in winter-run population, as observed in other salmonids (Garant *et al.* 2003).

Effects of different types of artificial propagation on N_e

Given the poor reproductive success of H_{trad} (Araki *et al.* 2006), the negative effect on N_h may not be surprising. However, in Su94–97 and Wi91, in which H_{trad} dominated, the variance in reproductive success of hatchery-born parents was high. Given that variance in reproductive success is the primary cause of $N_e/N < 1$, this result clearly suggests that H_{trad} decreased the effective size of the natural populations. Furthermore, the variance in reproductive success of wild-born fish was also increased in some of these run years. The cause of this increase is unclear, but one possible explanation is an indirect effect of H_{trad} on the natural populations. Under this scenario, variance in reproductive success is increased not only because of poor reproduction by the H_{trad} parents, but also by reduced reproductive success of wild-born parents that mated with the H_{trad} parents. Indeed, such a negative interaction was indicated between wild-born and $H_{\rm trad}$ parents (but not between wild-born and H_{supp} parents, Araki *et al.* 2006). If this is the case, the presence of H_{trad} has a direct impact on N_{ρ} in the natural population, and the removal of H_{trad} fish should increase N_e in subsequent generations, as expected in some situations (Ryman & Laikre 1991).

Similarities between wild-born and H_{supp} parents in reproductive success (Fig. 3, see also Araki *et al.* 2006) indicate that supplementation hatchery-born fish may actually support the natural populations. \hat{N}_e in the winter run generation-2 (Wi95–98) was 37% larger than generation-1 (Wi91–94, Table 2), and N_b was gradually increasing in winter run during 1995–98 (Fig. 2). These favourable results for conservation purposes in the winter run population might or might not be caused by the supplementation program, but we certainly found no sign of a negative effect of the program on the effective size of the natural population.

One caveat is that all the \hat{N}_e in this study were estimated based on direct information only from the anadromous offspring, and do not include nonanadromous offspring. Given that the frequent reproductive interaction in *Oncorhynchus mykiss*, production of nonanadromous offspring by anadromous parents is also likely to occur. If so, overall \hat{N}_e (including both anadromous and nonanadromous offspring) can be different when different types of parents (wild-born, H_{trad} , and H_{supp}) reproduce anadromous and nonanadromous offspring differently, as suggested in brown trout (*Salmo trutta* L., Hansen *et al.* 2000).

N_o/N ratio

The estimated \hat{N}_e/\hat{N} in this study (0.2–0.4) were not only between the ranges of estimates reported in this species in previous studies (0.1–0.3, Heath *et al.* 2002; 0.5–0.7, Ardren & Kapuscinski 2003), but also within the range of N_e/N that is suggested both theoretically and empirically for the single-generation N_e/N in various mating systems (0.3–0.5; see Nunney 1993, 1995; Frankham 1995b; Waples 2004 for reviews). The dominant effect of variance in reproductive success on reducing N_e/N was also consistent with the other studies on salmonids (Ardren & Kapuscinski 2003; Hedrick 2005b), suggesting that variance in reproductive success truly is the key factor to reduce N_e/N in salmon populations.

The extent to which variance in reproductive success plays an important role in determining N_e/N in other organisms remains largely unknown. However, recent studies indicate that its role can be substantial. For example, Turner *et al.* (2002) showed that variance in reproductive success is largely responsible for very small \hat{N}_e/\hat{N} (= 0.001) in red drum (*Sciaenops ocellatus*), whereas Matocq (2004) showed a moderately large \hat{N}_e/\hat{N} (= 0.48) in a woodrat (*Neotoma macrotis*) population that has low variance in reproductive success ($V_k \sim \bar{k}$) despite their population structure of harem polygyny.

Genetic compensation between life-history forms

In the Hood River steelhead, it appears that nonanadromous fish play an important role in stabilizing N_b and maintaining a high $N_{\rm e}/N_{\rm e}$ despite the variable census sizes of anadromous adults. The negative correlation between anadromous run size and inferred reproductive contribution of nonanadromous parents (Fig. 5) is most likely caused by a genetic compensation between life-history forms, in which a stable population of nonanadromous parents contributes a relatively constant number of offspring to the anadromous population every year. Another potential explanation for the relationship we observed is an increased competitive ability of nonanadromous parents in the spawning ground when the number of anadromous competitors is small. In this case, an increase in the absolute (rather than relative) contribution of nonanadromous parents to the anadromous populations is expected when anadromous run size is small. We did not detect any statistical evidence of negative correlation between the anadromous run size and the (inferred) absolute contribution of nonanadromous parents from our data (two-tailed *t*-test, P > 0.40), whereas the correlations tended to be negative (r = -0.49 to -0.03). This result indicates that the genetic compensation between life-history forms is primarily due to stable numbers of contributing nonanadromous parents, although a possibility of the increased competitive ability of nonanadromous parents remains.

A negative correlation between N_e/N and N has been reported in Atlantic salmon (Fraser et al. 2006), an insect (Pray et al. 1996), and a plant (Husband & Barret 1992), suggesting that genetic compensation is a common mechanism for the maintenance of genetic diversity. Given that many organisms have different life-history forms with different reproductive strategies within species (Clutton-Brock 1988), genetic compensation between life-history forms might also be common. In Atlantic salmon, for instance, a potentially important role of sexually matured parr on N_{e} is suggested (Jones & Hutchings 2002). As a practical consequence, conservation programmes that focus only on a part of life-history forms may not be sufficient to maintain genetic diversity throughout the entire populations. Our results emphasize the need to support the full range of diversity, including life-history polymorphisms.

Acknowledgements

We thank E. Olsen for technical advices, K. Kostow for arranging initial funding for the work, and D. Lytle, M. M. Hansen and two anonymous reviewers for useful comments. This research was funded by contracts to MSB from the Bonneville Power Administration and the Oregon Department of Fisheries and Wildlife.

References

- Anderson EC (2001) *Monte Carlo methods for inference in population genetic models.* PhD Dissertation, University of Washington, Seattle.
- Araki H, Blouin MS (2005) Unbiased estimation of relative reproductive success of different groups: evaluation and correction of bias caused by parentage assignment errors. *Molecular Ecology*, 14, 4097–4109.
- Araki H, Ardren WR, Olsen E, Cooper B, Blouin MS (2006) Reproductive success of captive-bred steelhead trout in the wild: Evaluation of three hatchery programs in the Hood River. *Conservation Biology*, published online, DOI: 10.1111/j.1523-1739.2006.00564.x.
- Ardren WR, Kapuscinski AR (2003) Demographic and genetic estimates of effective population size (*Ne*) reveals genetic compensation in steelhead trout. *Molecular Ecology*, **12**, 35–49.
- Ballou JD, Foose TJ (1996) Demographic and genetic management of captive populations. In: Wild Mammals in Captivity (eds Kleinman DG, Allen ME, Thompson KV, Lumpkin S), pp. 263– 283. The University of Chicago Press, Chicago.
- Caballero A (1994) Developments in the prediction of effective population size. *Heredity*, **73** (6), 657–679.
- Clutton-Brock TH (1988) Reproductive Success: Studies of Individual Variation in Contrasting Breeding Systems. University of Chicago Press, Chicago.
- Crow JF, Kimura M (1970) An Introduction to Population Genetics Theory. Harper & Row, New York.
- Crow JF, Morton NE (1955) Measurement of gene frequency drift in small populations. *Evolution*, **9**, 202–214.
- Cuenco ML, Barkman TWH, Mundy PR (1993) The Use of Supplementation to Aid in Natural Stock Restoration. In: *Genetic Conservation of Salmonid Fishes* (eds Cloud JG, Thorgaard GH), pp. 269–294. Plenum Press, New York.

- Duchesne P, Bernatchez L (2002) An analytical investigation of the dynamics of inbreeding in multi-generation supportive breeding. *Conservation Genetics*, 3, 45–58.
- Frankham R (1995a) Conservation genetics. Annual Review of Genetics, 29, 305–327.
- Frankham R (1995b) Effective population size/adult population size ratios in wildlife: a review. *Genetical Research*, **66**, 95–107.
- Frankham R, Briscoe DA, Ballou JD (2002) Introduction to Conservation Genetics. Cambridge University Press, Cambridge, UK.
- Fraser DJ, Jones MW, McParland TL, Hutchings JA (2006) Loss of historical immigration and the unsuccessful rehabilitation of extirpated salmon populations. *Conservation Genetics*, online early DOI 10.1007/s10592-006-9188-8.
- Garant D, Dodson JJ, Bernatchez L (2001) A genetic evaluation of mating system and determinants of individual reproductive success in Atlantic salmon (*Salmo salar* L.). *The Journal of Heredity*, 92, 137–145.
- Garant D, Dodson JJ, Bernatchez L (2003) Differential reproductive success and heritability of alternative reproductive tactics in wild Atlantic salmon (*Salmo salar L.*). *Evolution*, **57**, 1133–1141.
- Garcia-Vazquez E, Moran P, Martinez JL *et al.* (2001) Alternative mating strategies in Atlantic salmon and brown trout. *The Journal of Heredity*, **92**, 146–149.
- Gautschi B, Muller JP, Schmid B, Shykoff JA (2003) Effective number of breeders and maintenance of genetic diversity in the captive bearded vulture population. *Heredity*, **91**, 9–16.
- Hansen MM, Ruzzante DE, nielsen EE, Mensberg KL (2000) Microsatellite and mitochondrial DNA polymorphism reveals life-history dependent interbreeding between hatchery and wild brown trout (*Salmo trutta* L.). *Molecular Ecology*, **9**, 583–594.
- Hauser L, Adcock GJ, Smith PJ, Ramirez JH, Carvalho GR (2002) Loss of microsatellite diversity and low effective population size in an overexploited population of New Zealand snapper (*Pagrus auratus*). *Proceedings of the National Academy of Sciences of the USA*, **99**, 11742–11747.
- Heath DD, Busch C, Kelly J, Atagi DY (2002) Temporal change in genetic structure and effective population size in steelhead trout (Oncorhynchus mykiss). Molecular Ecology, 11, 197–214.
- Hedrick PW (2005a) *Genetics of Populations*, 3rd edn. Jones and Bartlett Publishers, Boston.
- Hedrick PW (2005b) Large variance in reproductive success and the *Ne/N* ratio. *Evolution*, **59**, 1596–1599.
- Hedrick PW, Hedgecock D, Hamelberg S, Croci SJ (2000) The impact of supplementation in winter-run chinook salmon on effective population size. *The Journal of Heredity*, **91**, 112–116.
- Husband B, Barrett S (1992) Effective population size and genetic drift in tristylous *Eichhornia paniculata* (Pontederiaceae). *Evolution*, 46, 1875–1890.
- Jones MW, Hutchings JA (2002) Individual variation in Atlantic salmon fertilization success: Implications for effective population size. *Ecological Applications*, **12**, 184–193.
- Kalinowski ST, Waples RS (2002) Relationship of effective to census size in fluctuating populations. *Conservation Biology*, **16**, 129–136.
- Kostow KE (2004) Differences in juvenile phenotypes and survival between hatchery stocks and a natural population provide evidence for modified selection due to captive breeding. *Canadian Journal of Fisheries and Aquatic Sciences*, **61**, 577–589.
- Leber KM (2004) Stock Enhancement and Sea Ranching: Developments, Pitfalls and Opportunities, 2nd edn. Blackwell Publications, Oxford, UK.

© 2006 The Authors

Journal compilation © 2006 Blackwell Publishing Ltd

- Marshall TC, Slate J, Kruuk LE, Pemberton JM (1998) Statistical confidence for likelihood-based paternity inference in natural populations. *Molecular Ecology*, 7, 639–655.
- Martinez JL, Moran P, Perez J *et al.* (2000) Multiple paternity increases effective size of southern Atlantic salmon populations. *Molecular Ecology*, **9**, 293–298.
- Matocq MD (2004) Reproductive success and effective population size in woodrats (*Neotoma macrotis*). *Molecular Ecology*, **13**, 1635–1642.
- Muñoz RC, Warner RR (2003) A new version of the size-advantage hypothesis for sex change: Incorporating sperm competition and size-fecundity skew. *American Naturalist*, **161**, 749–761.
- Nelson RJ, Beacham TD, Small MP (1998) Microsatellite analysis of the population structure of a Vancouver island sockeye salmon (*Oncorhynchus nerka*) stock complex using nondenaturing gel electrophoresis. *Molecular Marine Biology and Biotechnology*, 7, 312–319.
- Nunney L (1993) The influence of mating system and overlapping generations on effective population size. *Evolution*, **47**, 1329–1341.
- Nunney L (1995) Measuring the ratio of effective population size to adult numbers using genetic and ecological data. *Evolution*, 49, 389–392.
- Olney PJS, Mace GM, Feistner A (1994) *Creative Conservation: Interactive Management of Wild and Captive Animals*, 1st edn. Chapman & Hall, London.
- Olsen EA (2003) Hood River and Pelton ladder evaluation studies. In: Annual Report 2000–01 of the Oregon Department of Fish and Wildlife. Oregon Department of Fish and Wildlife, Portland.
- Pray LA, Goodnight CJ, Stevens L, Schwartz JM, Yan G (1996) The effect of population size on effective population size: an empirical study in the red flour beetle *Tribolium castaneum*. *Genetical Research*, 68, 151–155.
- Ryman N, Laikre L (1991) Effects of supportive breeding on the genetically effective population size. *Conservation Biology*, 5, 325–329.
- Salmon Recovery Science Review Panel (2004) Report for meeting held August 30-September 2, 2004, Northwest fisheries science center, National marine fisheries service, Seattle, Washington. Available from http://www.nwfsc.noaa.gov/trt/ rsrpreportsept30_2004b.pdf.
- Scribner KT, Arntzen JW, Burke T (1997) Effective number of breeding adults in *Bufo bufo* estimated from age-specific variation at minisatellite loci. *Molecular Ecology*, 6, 701–712.
- Sokal RR, Rohlf FJ (1995) Biometry: the Principles and Practice of Statistics in Biological Research, 3rd edn. W.H. Freeman, New York.
- Storz JF, Ramakrishnan U, Alberts SC (2002) Genetic effective size of a wild primate population: influence of current and historical demography. *Evolution*, 56, 817–829.
- Turner TF, Wares JP, Gold JR (2002) Genetic effective size is three orders of magnitude smaller than adult census size in an abundant, Estuarine-dependent marine fish (*Sciaenops ocellatus*). *Genetics*, **162**, 1329–1339.
- Wade MJ, Shuster SM (2004) Sexual selection: Harem size and the variance in male reproductive success. *The American Naturalist*, 164, E83–E89.
- Wang J, Ryman N (2001) Genetic effects of multiple generations of supportive breeding. *Conservation Biology*, **15**, 1619–1631.
- Waples RS (2002a) Effective size of fluctuating salmon populations. *Genetics*, 161, 783–791.
- Waples RS (2002b) Evaluating the effect of stage-specific survivorship on the *N*(*e*) /*N* ratio. *Molecular Ecology*, **11**, 1029–1037.

- Waples RS (2004) Salmonid insights into effective population size. In: *Evolution Illuminated: Salmon and Their Relatives* (eds Hendry AP, Stearns SC), pp. 295–314. Oxford University Press, Oxford, UK.
- Waples RS (2006a) Seed banks, salmon, and sleeping genes: effective population size in semelparous, age-structured species with fluctuating abundance. *American Naturalist*, **167**, 118–135.
- Waples RS (2006b) A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. *Conservation Genetics*, **7**, 167–184.
- Waples RS, Do C (1994) Genetic risk associated with supplementation of Pacific salmonids: captive broodstock programs. *Canadian Journal of Fisheries and Aquatic Sciences*, **51** (Suppl. 1), 310–329.
- Zimmerman CE, Reeves GH (2000) Population structure of sympatric anadromous and nonanadromous *Oncorhynchus mykiss*: evidence from spawning surveys and otolith

microchemistry. *Canadian Journal of Fisheries and Aquatic Sciences*, **57**, 2152–2162.

Hitoshi Araki's current research focuses on evolutionary and conservation biology of salmonids and population genetics on plant-pathogen interaction. Robin Waples is interested in developing and applying population genetic principles to realworld problems in ecology, conservation, and management. Mike Blouin's laboratory in Oregon State University focuses on the causes and consequences of genetic structuring, and on applications of methods for parentage analysis in natural populations. The research interests of the authors also include molecular evolution of gene systems, the evolutionary dynamics of genotypeenvironment interaction, and local adaptation. We are working on a variety of taxa including fish, amphibian, parasites, and plants.